

MRCK α , Active

Recombinant human protein expressed in Sf9 cells

Catalog # C27-11G-10

Lot # N331-2

Product Description

Recombinant human MRCK α (1-473) was expressed by baculovirus in Sf9 insect cells using an N-terminal GST tag. The gene accession number is [NM_003607](#).

Gene Aliases

CDC42BPA, MRCK, PK428, FLJ23347, KIAA0451, DKFZp686L1738, DKFZp686P1738

Concentration

0.1 $\mu\text{g}/\mu\text{l}$

Formulation

Recombinant protein stored in 50mM Tris-HCl, pH 7.5, 150mM NaCl, 10mM glutathione, 0.1mM EDTA, 0.25mM DTT, 0.1mM PMSF, 25% glycerol.

Storage, Shipping and Stability

Store product at -70°C . For optimal storage, aliquot target into smaller quantities after centrifugation and store at recommended temperature. For most favorable performance, avoid repeated handling and multiple freeze/thaw cycles. Stability is 1yr at -70°C from date of shipment. Product shipped on dry ice.

Scientific Background

Myotonic Dystrophy Kinase-Related Cdc42-Binding Kinase α (MRCK α) is a Cdc42/Rac/Rho interactive/binding serine/threonine kinase with multiple functional domains (1). MRCK are effectors of RhoA and Cdc42, respectively, for actin reorganization. MRCK α is a critical regulator of signal transduction pathways in eukaryotic cells that are known principally for their role in regulating the cytoskeleton, and they do so by recruiting a variety of downstream effector proteins (2).

References

- Ivan, T. et al: Genomic organization of human myotonic dystrophy kinase-related Cdc42-binding kinase α reveals multiple alternative splicing and functional diversity. *Gene*; 2003; 304:107-115.
- Ivan, T. et al: Phosphorylation of a Novel Myosin Binding Subunit of Protein Phosphatase 1 Reveals a Conserved Mechanism in the Regulation of Actin Cytoskeleton. *J. Biol. Chem.*, 2001; 276 (24): 21209-21216.

Purity

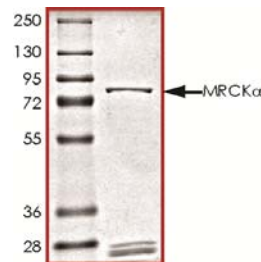
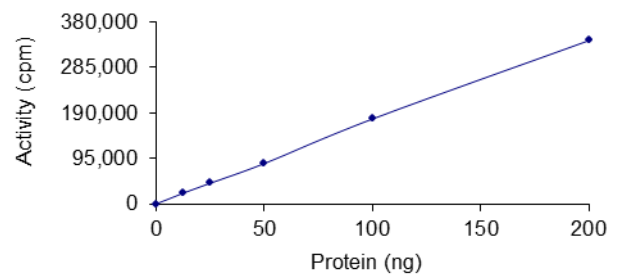


Figure 1. SDS-PAGE gel image

The purity of MRCK α was determined to be **>70%** by densitometry, approx. MW **~73kDa**.

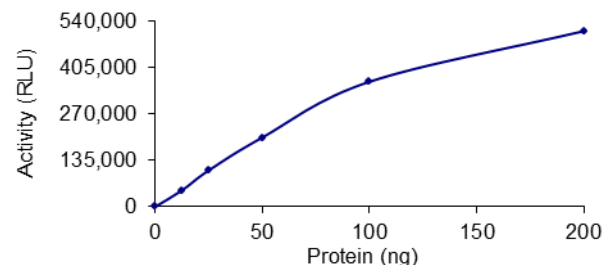
Specific Activity

Figure 2. Radiometric Assay Data



The specific activity of MRCK α was determined to be **86 nmol/min/mg** as per activity assay protocol. (For Radiometric Assay Protocol on this product please see pg. 2)

Figure 3. ADP-Glo™ Assay Data



The specific activity of MRCK α was determined to be **78 nmol/min/mg** as per activity assay protocol. (For ADP-Glo™ Assay Protocol on this product please see pg. 3)

Activity Assay Protocol

Reaction Components

Active Kinase (Catalog #: C27-11G)

Active MRCK α (0.1 μ g/ μ l) diluted with Kinase Dilution Buffer (Catalog #: K23-09) and assayed as outlined in sample activity plot. (Note: these are suggested working dilutions and it is recommended that the researcher perform a serial dilution of Active MRCK α for optimal results).

Kinase Dilution Buffer (Catalog #: K23-09)

Kinase Assay Buffer I (Catalog #: K01-09) diluted at a 1:4 ratio (5X dilution) with 50ng/ μ l BSA solution.

Kinase Assay Buffer (Catalog #: K01-09)

Buffer components: 25mM MOPS, pH 7.2, 12.5mM β -glycerol-phosphate, 25mM MgCl₂, 5mM EGTA, 2mM EDTA. Add 0.25mM DTT to Kinase Assay Buffer prior to use.

[³³P]-ATP Assay Cocktail

Prepare 250 μ M [³³P]-ATP Assay Cocktail in a designated radioactive working area by adding the following components: 150 μ l of 10mM ATP Stock Solution (Catalog #: A50-09), 100 μ l [³³P]-ATP (1mCi/100 μ l), 5.75ml of Kinase Assay Buffer (Catalog #: K01-09). Store 1ml aliquots at -20°C.

10mM ATP Stock Solution (Catalog #: A50-09)

Prepare ATP stock solution by dissolving 55mg of ATP in 10ml of Kinase Assay Buffer (Catalog #: K01-09). Store 200 μ l aliquots at -20°C.

Substrate (Catalog #: S05-58)

S6K synthetic peptide substrate (KRRRLASLR) diluted in distilled H₂O to a final concentration of 1mg/ml.

Assay Protocol

- Step 1. Thaw [³³P]-ATP Assay Cocktail in shielded container in a designated radioactive working area.
- Step 2. Thaw the Active MRCK α , Kinase Assay Buffer, Substrate and Enzyme Dilution Buffer on ice.
- Step 3. In a pre-cooled microfuge tube, add the following reaction components bringing the initial reaction volume up to 20 μ l:
 - Component 1. 10 μ l of diluted Active MRCK α (Catalog #C27-11G)
 - Component 2. 5 μ l of 1mg/ml stock solution of substrate (Catalog #S05-58)
 - Component 3. 5 μ l distilled H₂O (4°C)
- Step 4. Set up the blank control as outlined in step 3, excluding the addition of the substrate. Replace the substrate with an equal volume of distilled H₂O.
- Step 5. Initiate the reaction by the addition of 5 μ l [³³P]-ATP Assay Cocktail bringing the final volume up to 25 μ l and incubate the mixture in a water bath at 30°C for 15 minutes.
- Step 6. After the 15 minute incubation period, terminate the reaction by spotting 20 μ l of the reaction mixture onto individual pre-cut strips of phosphocellulose P81 paper.
- Step 7. Air dry the pre-cut P81 strip and sequentially wash in a 1% phosphoric acid solution (dilute 10ml of phosphoric acid and make a 1L solution with distilled H₂O) with constant gentle stirring. It is recommended that the strips be washed a total of 3 intervals for approximately 10 minutes each.
- Step 8. Count the radioactivity on the P81 paper in the presence of scintillation fluid in a scintillation counter.
- Step 9. Determine the corrected cpm by removing the blank control value (see Step 4) for each sample and calculate the kinase specific activity as outlined below.

Calculation of [³³P]-ATP Specific Activity (SA) (cpm/pmol)

Specific activity (SA) = cpm for 5 μ l [³³P]-ATP / pmoles of ATP (in 5 μ l of a 250 μ M ATP stock solution, i.e., 1,250 pmoles)

Kinase Specific Activity (SA) (pmol/min/ μ g or nmol/min/mg)

Corrected cpm from reaction / [(SA of ³³P-ATP in cpm/pmol)*(Reaction time in min)*(Enzyme amount in μ g or mg)]*[(Reaction Volume) / (Spot Volume)]

ADP-Glo™ Activity Assay Protocol

Reaction Components

MRCK α Kinase Enzyme System (Promega, Catalog #:V5710)

MRCK α , Active, 10 μ g (0.1 μ g/ μ l)
S6K Substrate, 1ml (1mg/ml)
Reaction Buffer A (5X), 1.5ml
DTT solution (0.1M), 25 μ l

ADP-Glo™ Kinase Assay Kit (Promega, Catalog #: V9101)

Ultra Pure ATP, 10 mM (0.5ml)
ADP, 10 mM (0.5ml)
ADP-Glo™ Reagent (5ml)
Kinase Detection Buffer (10ml)
Kinase Detection Substrate (Lyophilized)

Reaction Buffer A (5X)

200mM Tris-HCl, pH 7.5, 100mM MgCl₂ and 0.5 μ g/ μ l BSA.

Assay Protocol

The MRCK α assay is performed using the MRCK α Kinase Enzyme System (Promega; Catalog #: V5710) and ADP-Glo™ Kinase Assay kit (Promega; Catalog #: V9101). The MRCK α reaction utilizes ATP and generates ADP. Then the ADP-Glo™ Reagent is added to simultaneously terminate the kinase reaction and deplete the remaining ATP. Finally, the Kinase Detection Reagent is added to convert ADP to ATP and the newly synthesized ATP is converted to light using the luciferase/luciferin reaction. For more detailed protocol regarding the *ADP-Glo™ Kinase Assay*, see the Technical Manual #TM313, available at www.promega.com/tbs/tm313/tm313.html.

- Step 1. Thaw the ADP-Glo™ Reagents at ambient temperature. Then prepare Kinase Detection Reagent by mixing Kinase Detection Buffer with the Lyophilized Kinase Detection Substrate. Set aside.
- Step 2. Thaw the components of MRCK α Enzyme System, ADP and ATP on ice.
- Step 3. Prepare 1ml of 2X Buffer by combining 400 μ l Reaction Buffer A, 1 μ l DTT and 599 μ l of dH₂O.
- Step 4. Prepare 1ml of 250 μ M ATP Assay Solution by adding 25 μ l ATP solution (10mM) to 500 μ l of 2X Buffer and 475 μ l of dH₂O.
- Step 5. Prepare diluted MRCK α in 1X Buffer (diluted from 2X buffer) as outlined in sample activity plot. (Note: these are suggested working dilutions and it is recommended that the researcher perform a serial dilution of Active MRCK α for optimal results).
- Step 6. In a white 96-well plate (Corning Cat # 3912), add the following reaction components bringing the initial reaction volume up to 20 μ l:

Component 1.	10 μ l of diluted Active MRCK α
Component 2.	5 μ l of 1mg/ml stock solution of substrate
Component 3.	5 μ l of 2X Buffer
- Step 7. Set up the blank control as outlined in step 6, excluding the addition of the substrate. Replace the substrate with an equal volume of distilled H₂O.
- Step 8. At the same time as the MRCK α kinase reaction, set up an ATP to ADP conversion curve at 50 μ M ATP/ADP range as described in the *ADP-Glo™ Kinase Assay* Technical Manual #TM313.
- Step 9. Initiate the MRCK α reactions by the addition of 5 μ l of 250 μ M ATP Assay Solution thereby bringing the final volume up to 25 μ l. Shake the plate and incubate the reaction mixture at 30°C for 15 minutes.
- Step 10. Terminate the reaction and deplete the remaining ATP by adding 25 μ l of ADP-Glo™ Reagent. Shake the 96-well plate and then incubate the reaction mixture for another 40 minute at ambient temperature.
- Step 11. Add 50 μ l of the Kinase Detection Reagent, shake the plate and then incubate the reaction mixture for another 30 minute at ambient temperature.
- Step 12. Read the 96-well reaction plate using the Kinase-Glo™ Luminescence Protocol on a GloMax® Microplate Luminometer (Promega; Cat # E6501).
- Step 13. Using the conversion curve, determine the amount of ADP produced (nmol) in the presence (step 6) and absence of substrate (Step 7) and calculate the kinase specific activity as outlined below. For a detailed protocol of how to determine nmols from RLUs, see ADP-Glo™ Applications Database at <http://www.promega.com/applications/cellularanalysis/cellsignaling.htm>

Kinase Specific Activity (SA) (nmol/min/mg)

(ADP (step 6) – ADP (Step 7)) in nmol) / (Reaction time in min)*(Enzyme amount in mg)